

**Escherichioses Screen & Diff Real-™**  
Real Time PCR Kit for qualitative detection and  
differentiation of diarrheagenic  
E.coli (EPEC, ETEC, EIEC, EHEC, and EA<sub>g</sub>EC)

for use with RotorGene™ 3000/6000 (Corbett Research),  
SmartCycler® (Cepheid), iQ iCycler™ and iQ5™ (Biorad),  
MX3000P® and MX3005P® (Stratagene),  
Applied Biosystems® 7300/7500 Real Time PCR Systems (Applied)

**REF** B62-50FRT

**VER** 20.05.2011

GentaUR Molecular Products  
Voortstraat 49  
1910 Kampenhout, Belgium

## Escherichioses Screen & Diff Real - TM

### INTENDED USE

The **Escherichioses Screen & Diff Real - TM** is a Real-Time PCR test for test for qualitative detection and differentiation of diarrheagenic E.coli (EPEC, ETEC, EIEC, EHEC, and EA<sub>g</sub>EC) DNA in environmental compartments and clinical material by using real-time hybridization-fluorescence detection. DNA is extracted from samples, amplified using real time amplification with fluorescent reporter dye probes specific for E.Coli (EIEC) and Internal Control IC. Test contains an (IC) which serves as an amplification control for each individually processed specimen and to identify possible reaction inhibition.

### MATERIALS PROVIDED

- **PCR-mix-1 EIEC/EHEC**, 0,6 ml;
- **PCR-mix-1 EPEC/ETEC/EA<sub>g</sub>EC**, 0,6 ml;
- **PCR-mix-2-Flu**, 2 x 0,3 ml;
- **TaqF Polymerase**, 2 x 0,03 ml;
- **Positive Control DNA EIEC / EHEC / STI (C+ EIEC / EHEC / STI)**, 0,1 ml;
- **Positive Control DNA EPEC / ETEC / EA<sub>g</sub>EC (C+ EPEC / ETEC / EA<sub>g</sub>EC)**, 0,1 ml;
- **Negative Control C-**, 1,2 ml;\*
- **Internal Control IC**, 1,0 ml;\*\*
- **DNA-buffer**, 0,5 ml;

Contains reagents for 55 tests.

\* *must be used in the isolation procedure as Negative Control of Extraction.*

\*\* *add 10 µl of Internal Control IC during the DNA purification procedure directly to the sample/lysis mixture*

### MATERIALS REQUIRED BUT NOT PROVIDED

- Real Time Thermalcycler
- Workstation
- Pipettes (adjustable)
- Sterile tips with filters
- Tube racks
- DNA extraction kit

### WARNINGS AND PRECAUTIONS

1. Wear disposable gloves, laboratory coats and eye protection when handling specimens and reagents. Thoroughly wash hands afterward.
2. Do not pipette by mouth.
3. Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
4. Do not use a kit after its expiration date.
5. Dispose of all specimens and unused reagents in accordance with local regulations.
6. Biosafety Level 2 should be used for materials that contain or are suspected of containing infectious agents.
7. Clean and disinfect all spills of specimens or reagents using a disinfectant such as 0,5% sodium hypochlorite, or other suitable disinfectant.
8. Avoid contact of specimens and reagents with the skin, eyes and mucous membranes. If these solutions come into contact, rinse immediately with water and seek medical advice immediately.
9. Material Safety Data Sheets (MSDS) are available on request.
10. Use of this product should be limited to personnel trained in the techniques of amplification.
11. Workflow in the laboratory must proceed in a uni-directional manner, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where you performed previous step.

### STORAGE INSTRUCTIONS

**Escherichioses Screen & Diff Real - TM** must be stored at -20°C.

The complete kit can be shipped at 2-8°C but should be stored at -20°C immediately on receipt.

### STABILITY

**Escherichioses Screen & Diff Real - TM** Test is stable up to the expiration date indicated on the kit label. The product will maintain performance through the control date printed on the label. Exposure to light, heat or humidity may affect the shelf life of some of the kit components and should be avoided. Repeated thawing and freezing of these reagents should be avoided, as this may reduce the sensitivity

## SAMPLE COLLECTION, STORAGE AND TRANSPORT

**Escherichioses Screen & Diff Real - TM** can analyze DNA extracted with **DNA-Sorb-B** from:

- *Liquid cultures*;
- *water*: centrifuge 10-20 ml for 10 min at maximum speed. Discard the supernatant and leave about 100 µl of solution for DNA extraction;
- *feces*:
  - Prepare 20% feces suspension by adding in 5 ml tube of 4ml of Saline Solution and 1,0 gr (approx. 1,0 ml) of feces. Vortex to get the homogeneous suspension and centrifuge for 5 min to 7000-12000g and using a micropipette with a plugged aerosol barrier tip transfer in a new sterile 1,5 ml tube 100 µl of the bacterial fraction (white-yellowish line between the sediment and the supernatant)
  - Add 800 µl of PBS or Saline Solution. Vortex to get the homogeneous suspension and centrifuge for 5 min to 7000-12000g. Remove and discard the supernatant
  - Resuspend the pellet in 0,3 ml of PBS or Saline Solution.

It is recommended to process samples immediately after collection. Store samples at 2–8 °C for no longer than 24 hours, or freeze at –20/80°C. Transportation of clinical specimens must comply with country, federal, state and local regulations for the transport of etiologic agents.

## DNA ISOLATION

The following kit is recommended:

- ⇒ **DNA/RNA-Prep** (Sacace, **REF K-2-9**);
- ⇒ **DNA-Sorb-B** (Sacace, **REF K-1-1/B**)

Please carry out RNA extraction according to the manufacture's instruction.

Add 10 µl of Internal Control during DNA isolation procedure directly to the sample/lysis mixture.

**PROTOCOL (Reaction volume 25 µl):**

Total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

- 1 Prepare required quantity of reaction tubes for samples and controls.
- 2 Prepare the reaction mix for required number of samples, including controls. Mix **PCR-mix-1 EIEC / EHEC / STI** with **PCR-mix-Flu** and **polymerase (TaqF)** as well as **PCR-mix-1 PEC / ETEC / EA<sub>g</sub>EC** with **PCR-mix-2-FRT** and **polymerase (TaqF)** (see Table). Vortex the tube, then centrifuge shortly.

Table. **SCHEME OF REACTION MIXTURE PREPARATION**

Reagent volume per 1 reaction (µl)	Reagent volume per specified number of reactions (µl)		
	10.00	5.00	0.50
Number of reactions	PCR-mix-1-FRT	PCR-mix-2-FRT	Polymerase (TaqF)
6	60	30	3.0
8	80	40	4.0
10	100	50	5.0
12	120	60	6.0
14	140	70	7.0
16	160	80	8.0
18	180	90	9.0
20	200	100	10.0
22	220	110	11.0
24	240	120	12.0
26	260	130	13.0
28	280	140	14.0
30	300	150	15.0
32	320	160	16.0

- 3 Vortex the tube, then centrifuge shortly. Add **15 µl** of prepared reaction mix into each appropriate tube.
- 4 Using tips with aerosol filter add **10 µl** of DNA samples obtained at the stage of DNA isolation and mix carefully by pipetting.

*N.B. If the DNA-Sorb isolation kit is used as a DNA extraction kit, re-centrifuge all the tubes with extracted DNA for 2 min at maximum speed (12000-16000 g) and take carefully supernatant. N.B. don't disturb the pellet, sorbent inhibit reaction*

- 5 Prepare for each panel 3 controls:
  - add **10 µl** of **DNA-buffer** to the tube labeled Negative Control Amplification (NCA);
  - add **10 µl** of **Positive Control DNA EIEC / EHEC / STI** to the tube labeled C+ *EIEC / EHEC / STI* (Positive Control of Amplification) for **PCR-mix-1 EIEC / EHEC / STI**;
  - add **10 µl** of **Positive Control DNA EPEC / ETEC / EA<sub>g</sub>EC** to the tube labeled C+ *EPEC / ETEC / EA<sub>g</sub>EC* (Positive Control of Amplification) for **PCR-mix-1 EPEC / ETEC / EA<sub>g</sub>EC**.

Create a temperature profile on your Real-time instrument as follows:

Stage	Rotor type instruments <sup>1</sup>				Plate type or modular instruments <sup>2</sup>			
	Temp, °C	Time	Fluorescence detection	Cycle repeats	Temp, °C	Time	Fluorescence detection	Cycle repeats
Hold	95	15 min	–	1	95	15 min	–	1
Cycling 2	95	10 s	–	45	95	10 s	–	45
	60	25 s	FAM(Green), JOE(Yellow)		60	30 s	FAM, JOE/HEX/Cy3	
	72	10 s	–		72	10 s	–	

<sup>1</sup> For example Rotor-Gene™ 3000/6000 (Corbett Research, Australia)

<sup>2</sup> For example, iQ5™/iQ iCycler™ (BioRad, USA); Mx3000P/Mx3005P™ (Stratagene, USA), Applied Biosystems® 7300/7500 Real Time PCR (Applied Biosystems), SmartCycler® (Cepheid)

## INSTRUMENT SETTINGS

*Escherichioses*” test settings for rotor-type instruments (Rotor-Gene 6000, Rotor-Gene Q etc.)

Channel	Threshold	More Settings/Outlier Removal	Slope Correct	Auto gain calibration channel settings
FAM/Green	0.05	10%	On	3-7 FL
JOE/Yellow	0.05	10%	On	3-7 FL
ROX/Orange	0.05	10%	On	5-10 FL

### Plate- or modular type instruments

For result analysis, set the threshold line at a level corresponding to 10–20% of the maximum fluorescence signal obtained for Pos C+ sample during the last amplification cycle.

### RESULTS ANALYSIS:

- The results are interpreted by the device software through the presence of crossing of fluorescence curve with the threshold line.

#### PCR-mix-1 *EIEC* / *EHEC* / *STI*:

Internal Control is detected on Fam/Green channel,  
*EHEC* is detected on the Joe/Yellow/HEX/Cy3 channel,  
*EIEC* on the Rox/Orange/TexasRed channel

#### PCR-mix-1 *EPEC* / *ETEC* / *EAgEC*:

*EAgEC* is detected on Fam/Green channel,  
*EPEC* is detected on the Joe/Yellow/HEX/Cy3 channel,  
*ETEC* on the Rox/Orange/TexasRed channel

Table. Interpretation of results for PCR-analysis

PCR-mix-1	Ct value in channel			Interpretation
	Fam/Green	Joe/Yellow/HEX/Cy3	Rox/Orange/TexasRed	
PCR-mix-1 <i>EIEC</i> / <i>EHEC</i> / <i>STI</i>	Pos (< 40)	Neg	Neg	<i>EIEC</i> and <i>EHEC</i> DNA is not detected
	Neg	Pos (< 40)	Neg	<i>EHEC</i> DNA is detected
	Neg	Neg	Pos (< 40)	<i>EIEC</i> DNA is detected
	Neg	Neg	Neg	invalid
PCR-mix-1 <i>EPEC</i> / <i>ETEC</i> / <i>EAgEC</i>	Pos (< 40)	Neg	Neg	<i>EAgEC</i> DNA is detected
	Neg	Pos	Neg	<i>EPEC</i> DNA is detected
	Neg	Neg	Pos (< 40)	<i>ETEC</i> DNA is detected
	Neg	Neg	Neg	<i>EPEC</i> / <i>ETEC</i> / <i>EAgEC</i> DNA are not detected

- The result of the analysis is considered reliable only if the results obtained for both Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct.

Table . Results for controls

PCR-mix-1	Control	Stage for control	Ct value in channel		
			Fam/Green	Joe/Yellow/HEX /Cy3	Rox/Orange /TexasRed
PCR-mix-1-FEP/FRT <i>EIEC</i> / <i>EHEC</i> / <i>STI</i>	C-	DNA extraction	Pos (< 40)	Neg (> 40)	Neg (> 40)
	NCA	Amplification	Neg (> 40)	Neg (> 40)	Neg (> 40)
	C+ <i>EIEC</i> / <i>EHEC</i> / <i>STI</i>	Amplification	Pos (< 40)	Pos (< 40)	Pos (< 40)
PCR-mix-1-FEP/FRT <i>EPEC</i> / <i>ETEC</i> / <i>EAgEC</i>	C-	DNA extraction	Neg (> 40)	Neg (> 40)	Neg (> 40)
	NCA	Amplification	Neg (> 40)	Neg (> 40)	Neg (> 40)
	C+ <i>EPEC</i> / <i>ETEC</i> / <i>EAgEC</i>	Amplification	Pos (< 40)	Pos (< 40)	Pos (< 40)

## TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

- If the signal for C<sup>-</sup> (except for C<sup>-</sup> in FAM channel for PCR-mix-1-FEP/FRT *EIEC* / *EHEC* / STI) and/or for NCA is less than the boundary value, analysis should be repeated starting from the DNA extraction stage.
- If no signal is detected for the positive controls of amplification, it may suggest that the programming of the temperature profile of the used instrument was incorrect, or that the configuration of the PCR reaction was incorrect, or that the storage conditions for kit components did not comply with the manufacturer's instruction, or that the reagent kit expired. Programming of the used instrument, storage conditions, and the expiration date of the reagents should be checked, and then PCR should be repeated.
- If a positive result (the fluorescence curve crosses the threshold line) is detected for a sample that has a fluorescence curve without the typical exponential growth phase (the curve is linear), this may suggest incorrect setting of the threshold line or incorrect calculation of baseline parameters. Such a result should not be considered as positive. Once the threshold line has been set correctly, PCR analysis of the sample should be repeated (if iCycler iQ or iQ5 instruments are used).

## PERFORMANCE CHARACTERISTICS

### Analytical specificity

The analytical specificity of **Escherichioses Screen & Diff Real - TM** PCR kit is ensured by selection of specific primers and probes as well as strict reaction conditions. The primers and probes were checked for possible homologies to all sequences deposited in gene banks by sequence comparison analysis. Nonspecific responses were absent during examination of human DNA as well as a DNA panel of the following microorganisms:

- *E.coli* strains: O157H7 No. 4, O157H7 No. 23, O157H7 No. 212, O157H7 No. 214, O157H7 No. 1330, O143, O124 No. 227, O144, O86 No. 990, O125 Carioni, O85, O61 No. 10167B/41, O59 No. 9095/41, No. 409 (O34), K12, 3912/41, Krym No. 56, O148H28 B7a, O6 No. 3091, 113/3, 675, O111 No. 153, O62 10524/41, O126 No. 611, M17, Krym No. 1274, 168/59, O57 8198/41, Krym No. 14169, O48, NCTC 9001.
- Strains of other microorganisms: *Salmonella enteritidis* S-6, *S.choleraesuis* 370, *S.typhimurium* 371, *S.dublin* 373, *S.typhi* C1, *S.abortusovis* 372, and *S.gallinarum-pullorum*; *Shigella flexneri* 851b; *Campylobacter fetus* ssp. *fetus* 25936 and *C.jejuni* ssp. *jejuni* 43435; *Klebsiella K 65* SW4; *Listeria monocytogenes* USHCH 19 and *L.monocytogenes* USHCH 52; *Proteus vulgaris* 115/98; *Pseudomonas aeruginosa* DH c1; *Staphylococcus aureus* 653 and *S. aureus* 29112; *Morganella morganii* 619 c 01; *Enterococcus faecalis* 356, 12 strains of *Yersinia enterocolitica*, and 6 strains of *Yersinia pseudotuberculosis*.

The specificity of diarrheagenic *E.coli* strains was confirmed by sequence analysis of the studied genome fragments.


### Analytical sensitivity

The kit **Escherichioses Screen & Diff Real - TM** allows to detect *E.Coli* DNA in 100% of the tests with a sensitivity of not less than 1000 copies/ml. The detection was carried out on the control standard and its dilutions by negative sample.

## EXPLANATION OF SYMBOLS

 REF Catalogue Number


 LOT Lot Number


 Expiration Date

  $\Sigma$  Contains reagents

 ! Caution!

 VER Version

 Manufacturer

 Temperature limitation

