Human soluble myosin heavy chain 1, sMHC-1 ELISA kit

Catalog No. E0754h

(96 tests)

Operating instruction

FOR RESEARCH USE ONLY; NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS!
PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING!

Intended use

This immunoassay kit allows for the specific measurement of human soluble myosin heavy chain 1, sMHC-1 concentrations in cell culture supernates, serum and plasma.

Introduction

Myosin heavy chains (MYHCs) are ubiquitous actin-based motor proteins that convert the chemical energy derived from hydrolysis of ATP into mechanical force that drives diverse motile processes, including cytokinesis, vesicular transport, and cellular locomotion, in eukaryotic cells. The MYHCs have been divided into 9 to 11 classes. Class II, or 'conventional,' MYHCs include the extensively studied group of sarcomeric MYHCs that self associate to form filaments and function enzymatically to promote contraction in striated (cardiac and skeletal) muscles. Muscle myosin is a heterohexamer consisting of 2 myosin heavy chains and 2 associated nonidentical pairs of myosin light chains. The 7 MYHC isoforms that predominate in mammalian skeletal muscles include 2 developmental isoforms, MYHC-embryonic (MYH3) and MYHC-perinatal (MYH8); 3 adult skeletal muscle isoforms, MYHC IIa (MYH2), MYHC IIb (MYH4), and MYHC IIx/d (MYH1); and MYHC-beta/slow (MYH7), which is also expressed in cardiac muscle. MYHC-extraocular (MYH13) is expressed primarily in extrinsic eye muscles.

Test principle

This assay employs the quantitative sandwich enzyme immunoassay technique. A antibody specific for sMHC-1 has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any sMHC-1 present is bound by the immobilized antibody. An enzyme-linked antibody specific for sMHC-1 is added to the wells. Following a wash to remove any unbound antibody-enzyme reagent, a substrate solution is added to the wells and color develops in proportion to the amount of sMHC-1 bound in the initial step. The color development is stopped and the intensity of the color is measured.

Materials and components

Reagent	Quantity
Assay plate	1
Standard	2
Sample Diluent	1 x 20ml
Assay Diluent A	1 x 10ml

Assay Diluent B	1 x 10ml
Detection Reagent A	1 x 120ul
Detection Reagent B	1 x 120ul
Wash Buffer	1 x 30ml
(25 x concentrate)	
Substrate	1 x 10ml
Stop Solution	1 x 10ml

Sample collection and storage

Cell culture supernates - Remove particulates by centrifugation and assay immediately or aliquot and store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at approximately 1000 x g. Remove serum and assay immediately or aliquot and store samples at -20° C.

Plasma - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000 x g at 2 - 8° C within 30 minutes of collection. Store samples at ≤ -20° C. Avoid repeated freeze-thaw cycles.

Note: Citrate plasma has not been validated for use in this assay.

Limitations of the procedure

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1. The kit should not be used beyond the expiration date on the kit label.

2. Do not mix or substitute reagents with those from other lots or sources.

3. If samples generate values higher than the highest standard, further dilute the samples with the Assay Diluent and repeat the assay. Any variation in standard diluent, operator, pipetting technique, washing technique,incubation time or temperature, and kit age can cause variation in binding.

4. This assay is designed to eliminate interference by soluble receptors, ligands, binding proteins, and other factors present in biological samples. Until all factors have been tested in the Quantikine Immunoassay, the possibility of interference cannot be excluded.

Reagent preparation

Bring all reagents to room temperature before use.

Wash Buffer - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Dilute 20 mL of Wash Buffer Concentrate into deionized or distilled water to prepare 500 ml of Wash Buffer.

Standard - Reconstitute the **Standard** with 1.0 mL of **Sample Diluent**. This reconstitution produces a stock solution of 100 ng/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making serial dilutions. The undiluted standard serves as the high standard (100 ng/mL). The **Sample Diluent** serves as the zero standard (0 ng/mL).

Detection Reagent A and B - Dilute to the working concentration specified on the vial label using **Assay Diluent A and B** (1:100), respectively.

Assay procedure

Allow all reagents to reach room temperature. Arrange and label required number of strips.

1. Prepare all reagents, working standards and samples as directed in the previous sections.

2. Add 100 uL of **Standard**, Control, or sample* per well. Cover with the adhesive strip. Incubate for 2 hours at 37° C.

3. Remove the liquid of each well, don't wash.

4. Add 100 uL of **Detection Reagent A** to each well. Incubate for 1 hour at 37°C. **Detection Reagent A** may appear cloudy. Warm to room temperature and mix gently until solution appears uniform.

5. Aspirate each well and wash, repeating the process three times for a total of three washes. Wash by filling each well with Wash Buffer (350 uL) using a squirt bottle, multi-channel pipette, manifold dispenser or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.

6. Add 100 uL of **Detection Reagent B** to each well. Cover with a new adhesive strip.Incubate for 1 hours at 37° C.

7. Repeat the aspiration/wash as in step 5.

8. Add 90 uL of **Substrate Solution** to each well. Incubate for 30 minutes at room temperature. Protect from light.

9. Add 50 uL of **Stop Solution** to each well. If color change does not appear uniform, gently tap the plate to ensure thorough mixing.

10. Determine the optical density of each well within 30 minutes, using a microplate reader set to 450 nm.

Specificity
This assay recognizes recombinant and natural human sMHC-1. No significant cross-reactivity or interference was observed.

Sensitivity
The minimum detectable dose of human sMHC-1 is typically less than 0.39 ng/mL.
The sensitivity of this assay, or Lower Limit of Detection (LLD) was defined as the lowest detectable concentration that could be differentiated from zero.

Detection Range
1.56-100 ng/mL. The assay range was estimated by calculating the coefficient of variation (CV) of each standard constructing five independent standard curves. The standard curve concentrations used for the ELISA's were 100 ng/mL, 50 ng/mL, 25 ng/mL, 12.5 ng/mL, 6.25 ng/mL, 3.12 ng/mL, 1.56 ng/mL.

Important Note:
1. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.
2. It is recommended that no more than 32 wells be used for each assay run if manual pipetting is used since pipetting of all standards, specimens and controls should be completed within 5 minutes. A full plate of 96 wells may be used if automated pipetting is available.
3. Duplication of all standards and specimens, although not required, is recommended.

4. When mixing or reconstituting protein solutions, always avoid foaming.
5. To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
6. To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.

Calculation of results

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the sMHC-1 concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

Storage of test kits and instrumentation

1. Unopened test kits should be stored at 2-8°C upon receipt and the microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air. The test kit may be used throughout the expiration date of the kit (six months from the date of manufacture). Refer to the package label for the expiration date.
2. Opened test kits will remain stable until the expiring date shown, provided it is stored as prescribed above.
3. A microtiter plate reader with a bandwidth of 10nm or less and an optical density range of 0-3 OD or greater at 450nm wavelength is acceptable for use in absorbance measurement.

Precaution

The Stop Solution suggested for use with this kit is an acid solution. Wear eye, hand, face, and clothing protection when using this material.