

User Bulletin

Viral RNA Extraction Miniprep System

For serum, plasma, body fluids, and cell culture supernatant.

Downstream Application

- * Reverse transcription
- * Northern blotting
- * PCR
- * RNase protection assay

Product Contents

Cat. No	GVR1001	GVR1002
Preps	50	250
RXV Buffer	35ml	190ml
WS Buffer	15ml	45ml x 2
RNA Carrier	1	1
RNase-free ddH ₂ O	1.5ml x 2	15ml
RNA Mini Column	50	250
Collection Tube	50	250
Elution Tube	50	250
Protocol	1	1

All buffers need to be mixed well before use.

Shipping & Storage

Viogene Viral RNA Extraction System is shipping and storage at ambient temperature up to 6 months.

If precipitate form by freezing temperature on any buffer, warm up at 37°C to redissolve.

Protocol

❖ **Please read the following notes before starting the procedures.**

- **WARNING**, strong acids and oxidants (like for instance bleach) should not be used together with RXV buffer (because this kind of reaction would produce cyanide)!!!

Important Notes

<Note>: Preheat RNase-free ddH₂O to 80°C.

-for GVR1001- Add 60 ml of ethanol (98-100%) to the WS Buffer bottle when first open the bottle.

-for GVR1002- Add 180 ml of ethanol (98-100%) to the WS Buffer bottle when first open the bottle.

1. Add RNA carrier to RXV Buffer.

Add 1 ml RXV Buffer to the RNA carrier tube, vortex to dissolve and transfer to the RXV Buffer bottle, store at 4°C.

2. Pipet 150 µl sample (serum, plasma, body fluids, and cell culture supernatant) into a 1.5 ml tube.

3. Add 570 µl of carrier added RXV Buffer to the sample, mix by vortexing.

Through mixing is required for sample lysis. If the sample volume is larger than 150 µl, increase the amount of RXV Buffer proportionally.

4. Incubate the vortexed sample at room temperature for 10 minutes.

5. Add 570 µl of ethanol (98-100%) to the sample, and mix by vortexing.

If the starting sample is larger than 150 µl, increase the amount of ethanol proportionally.

6. Place a Total RNA Column in a 2 ml Collection Tube, apply 650 µl of the ethanol added sample from step 5 to the Total RNA Column, close the cap, centrifuge at 6,000 x g (8,000 rpm) for 1 minute, and discard the filtrate.

If the solution remains above the membrane, centrifuge again at 13,000 rpm.

7. Repeat step 6 for rest of the sample.

8. Wash the column twice with 500 µl of ethanol added WS Buffer by centrifuging at full speed (13,000 rpm or 10,000 x g) for 1 minute, and discard the filtrate.

Add 60 ml (for GVR1001) or 180ml (for GVR1002) of ethanol (98-100%) to the WS Buffer bottle when first open the bottle.

9. Centrifuge at full speed for 3 minutes to remove traces of WS Buffer.

Residual ethanol may inhibit reverse transcriptase activity.

10. Transfer the column to a RNase-free 1.5 ml Elution Tube, add 50 µl of preheated (80°C) RNase-free ddH₂O, and centrifuge at full speed for 1 minute to elute RNA.

11. Store RNA at -70°C.