



Version 1.0909

**EpiQuik™ Total Histone H3
Quantification Kit (Fluorometric)**

Catalog No. P-3063

User Guide*

***Always use the most updated User
Guide included in your current order.**

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FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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INTRODUCTION

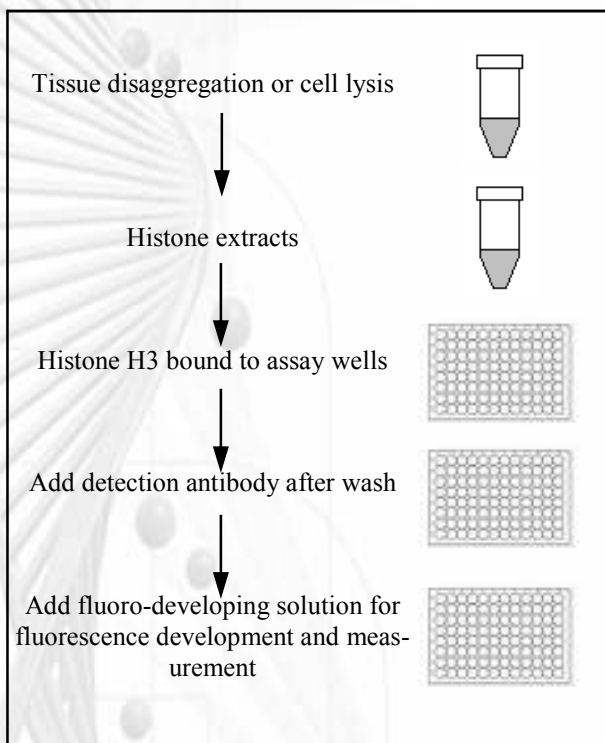
Histone H3 along with H2A, H2B and H4 is involved in the structure of chromatin in eukaryotic cells. Histone H3 can undergo several different types of epigenetic modifications that influence cellular processes such as transcription activation/inactivation, chromosome packaging and DNA damage repair. These modifications, including acetylation, phosphorylation, methylation, ubiquitination, and ADP-ribosylation, occur on the N-terminal tail domains of histone H3 through catalyzing of histone modifying enzymes, which results in remodeling of the nucleosome structure into an open conformation more accessible to transcription complexes. In most species, histone H3 is primarily acetylated at lysine 9, 14, 18, 23, and 56, and methylated at lysine 4, 9, 27, 36 and 79 and phosphorylated at ser10, ser28, Thr3 and Thr11, respectively. Thus, quantitative detection of various histone modifications would provide useful information for better understanding epigenetic regulation of cellular processes and for developing HMT-targeted drugs.

Epigentek has provided a series of kits used for quantifying all sites/degrees of histone H3 modification. For added convenience and more quantitative interpretation of the results with use of these kits, we provide here the *EpiQuik™* Total Histone H3 Quantification Kit (Fluorometric). This kit is designed for quantifying the level of histone H3 protein independent of its modified state and can also be used for normalizing the modified histone H3 content of samples when run in parallel with Epigentek histone modification quantification kit series. The kit has the following features:

- Quick and efficient procedure, which can be finished within 2.5 hours.
- Innovative fluorometric assay without need for radioactivity, electrophoresis, and chromatography.
- Specifically capturing histone H3 with the detection limit as low as 2 ng/well and detection range from 10 ng-1 μ g/well of histone extracts.
- The unmodified Histone H3 control is conveniently included for quantification of the amount of total histone H3.
- Strip microplate format makes the assay flexible: manual or high throughput.
- Simple, reliable, and consistent assay conditions.

PRINCIPLE AND PROCEDURE

The *EpiQuik*[™] Total Histone H3 Quantification Kit (Fluorometric) is designed for quantifying the level of histone H3 protein independent of its modified state. In an assay with this kit, the histone H3 protein is captured to the strip wells coated with anti-histone H3 antibody. The captured histone H3 can then be detected with a detection antibody followed by a fluorescent development reagent. The ratio of histone H3 is proportional to the intensity of fluorescence. The absolute amount histone H3 can be quantified by comparing to the standard control.



Schematic Procedure for Using the *EpiQuik*[™] Total Histone H3 Quantification Kit (Fluorometric)

PRODUCT USE INFORMATION

The *EpiQuik*[™] Total Histone H3 Quantification Kit (Fluorometric) is suitable for specifically measuring total histone H3 using human, mouse and rat samples including fresh and frozen tissues, cultured adherent and suspension cells.

The *EpiQuik*[™] Total Histone H3 Quantification Kit (Fluorometric) is for research use only and is not intended for diagnostic or therapeutic application.

Suitable lab coat, disposable gloves, and eye protection are required when working with the kit.

Epigentek guarantees the performance of all products in the manner described in our product instructions.

Epigentek reserves the right to change or modify any product to enhance its performance and design.

EpiQuik[™] is a trademark of Epigentek Group Inc.

KIT CONTENTS

Components	48 assays P-3063-48	96 assays P-3063-96
F1 (10X wash buffer)	10 ml	20 ml
F2 (antibody buffer)	6 ml	12 ml
F3 (detection antibody, 1 mg/ml)*	5 μ l	10 μ l
F4 (fluoro developer)*	12 μ l	24 μ l
F5 (fluoro enhancer)*	12 μ l	24 μ l
F6 (fluoro dilution)	4 ml	8 ml
Standard control (100 μ g/ml)*	10 μ l	20 μ l
8 well sample strips (with frame)	4	9
8 well standard control strips	2	3
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* For maximum recovery of the products, centrifuge the original vial prior to opening the cap.

SHIPPING AND STORAGE

Upon receipt, store **F3**, **F4**, and **standard control** at -20°C . Store **all other components** at 4°C away from light. The components of the kit should be stable for 6 months when stored properly.

Note: Check if buffers **F1** and **F2** contain salt precipitates before using. If so, warm (at room temperature or 37°C) and shake the buffers until the salts are re-dissolved.

MATERIALS REQUIRED BUT NOT SUPPLIED

Orbital shaker
Pipettes and pipette tips
Reagent reservoir
Fluorescence microplate reader
15 ml conical tube
1.5 ml microcentrifuge tubes

PROTOCOL

- Prepare histone extracts from cells/tissues treated or untreated by using your own successful method (acid extraction or high salt extraction).
 - For your convenience and best results, Epigentek offers the *EpiQuik*[™] Total Histone Extraction Kit (Cat # OP-0006) optimized for use in the *EpiQuik*[™] modified histone quantification series.
 - Preparation of histone extracts can also be performed using the attached procedure (See Appendix). Histone extracts can be used immediately or stored at -80°C for future use.
- Determine the number of strip wells required. Leave these strips in the plate frame (remaining unused strips can be placed back in the bag. Seal the bag tightly and store at 4°C). Dilute **F1** with distilled water (pH 7.2-7.5) at 1:9 ratio (1 ml of **F1** + 9 ml of distilled water).
- Add $50\ \mu\text{l}$ of **F2** into each well. For the sample, add 1-2 μg of the histone extract into the sample wells. For standard curve, dilute **standard control** with **F2** to 1 – 100 ng/ μl for 5-7 points (e.g., 1.5, 3, 6, 12, 25, 50, and 100 ng/ μl). Add 1 μl of **standard control** at the different concentrations into the standard well. For the blank, add no nuclear extracts or no standard control protein. Mix and cover the strip wells with Parafilm M and incubate at room temperature for 1-2 hours.
- Aspirate and wash the wells with 150 μl of **diluted F1** 3 times.
- Dilute **F3** (at 1:1000 ratio) to 1 $\mu\text{g}/\text{ml}$ with **F2**. Add 50 μl of diluted **F3** to each well and incubate at room temperature for 60 min on an orbital shaker (100 rpm).
- Aspirate and wash the wells with 150 μl of **diluted F1** 6 times.
- Prepare **fluoro-development solution** by adding 1 μl of **F4** and 1 μl of **F5** into each 400 μl of **F6**. Add 50 μl of **fluoro-development solution** into the wells and incubate at room temperature for 1-5 min away from light. The color in the standard wells containing the higher concentrations may turn slightly pink during this period. Measure and read fluorescence on fluorescence microplate reader at $530_{\text{EX}}/590_{\text{EM}}$ nm.

Note: If the strip well frame does not fit the fluorescence reader, transfer the solution to the standard 96-well microplate and read fluorescence at $530_{\text{EX}}/590_{\text{EM}}$ nm.

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8. Calculate % histone H3:

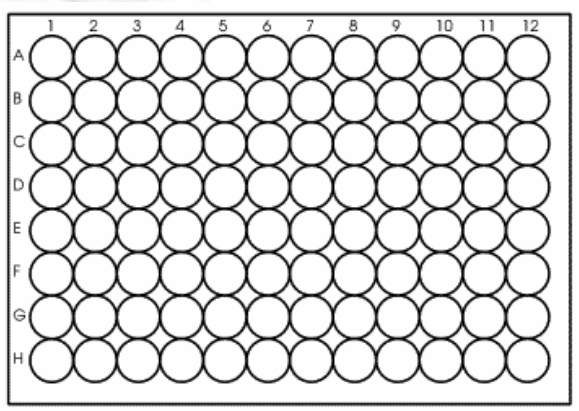
Plot delta RFU versus amount of **standard control** and determine the slope as delta RFU/ng.

Calculate the amount of Histone H3 using the following formula:

$$\text{Amount (ng/mg protein)} = \frac{\text{RFU (sample - blank)}}{\text{Protein } (\mu\text{g})^* \times \text{slope}} \times 1000$$

* Histone extract amount added into the sample well at step 3.

Plate Configuration



- **Strip 1-3 (for 96 assays) or strip 1-2 (for 48 assays):** standard wells (**Labeled as SC**); the standard curve can be generated with 5-8 concentration points (includes blank).
- Example amount of standard control/well: **A1:** 60 ng; **B1:** 30 ng; **C1:** 15 ng; **D1:** 7.5 ng; **E1:** 3.8 ng; **F1:** 1.9 ng; **G1:** 1 ng; **H1:** 0 ng.
- **Strip 4-12 (for 96 assays) or strip 3-6 (for 48 assays):** sample wells (**No label**).
- Each sample or standard point can be assayed in duplicates or triplicates.

Histone Extraction Protocol

1. For tissues (treated and untreated), weigh and cut the sample into a small piece (1-2 mm³) with a scalpel or scissors. Transfer tissue pieces to a Dounce homogenizer, add TEB buffer (PBS containing 0.5% Triton X 100, 2 mM PMSF and 0.02% NaN₃) at 200 mg/ml, and disaggregate tissue pieces by 50-60 strokes. Transfer homogenized mixture to a 15 ml conical tube and centrifuge at 3000 rpm for 5 min at 4°C. If total mixture volume is less than 2 ml, transfer mixture to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

For cells (treated and untreated). Harvest cells and pellet the cells by centrifugation at 1000 rpm for 5 min at 4°C. Resuspend cells in TEB buffer at 10⁷ cells/ml and lyse cells on ice for 10 min with gentle stirring. Centrifuge at 3000 rpm for 5 min at 4°C. If total volume is less than 2 ml, transfer cell lysates to a 2 ml vial and centrifuge at 10,000 rpm for 1 min at 4°C. Remove supernatant.

2. Resuspend cell/tissue pellet in 3 volumes (approx. 200 μ l/10⁷ cells or 200 mg tissues) of extraction buffer (0.5N HCl + 10% glycerol) and incubate on ice for 30 min.
3. Centrifuge at 12,000 rpm for 5 min at 4°C and remove the supernatant fraction to new vial.
4. Add 8 volumes (approx. 0.6 ml/10⁷ cells or 200 mg tissues) of acetone and leave at -20°C overnight.
5. Centrifuge at 12,000 rpm for 5 min and air-dry the pellet. Dissolve the pellet in distilled water (30-50 μ l/10⁷ cells or 200 mg tissues).
6. Quantify the protein concentration. Aliquot the extract and store the extract at -20°C or -80°C.

TROUBLESHOOTING

No Signal for Both the Standard Control and the Samples

Reagents are added incorrectly.

Check if reagents are added in order and if some steps of the procedure are omitted by mistake.

Incubation time and temperature is incorrect.

Ensure the incubation time and temperature described in the protocol are correctly followed.

No Signal or Very Weak Signal for Only the Standard Control

The amount of standard control is not added into “standard control wells” or is added insufficiently.

Ensure sufficient amount of control is properly added to the standard control well.

No Signal for Only the Sample

The protein sample is not properly extracted.

Ensure the procedure and reagents are correct for the nuclear protein extraction.

The protein amount is added into well insufficiently.

Ensure extract contains sufficient amounts of protein.

Protein extracts are incorrectly stored.

Ensure the nuclear extracts are stored at -20°C or -80°C .

High Background Present for the Blank

The well is not washed sufficiently.

Check if wash at each step is performed according to the protocol.

Contaminated by the standard control.

Ensure the well is not contaminated by adding the control protein or by using control protein contaminated tips.

Overdevelopment.

Decrease development time in Step 7.

ORDERING INFORMATION

Products	Size	Cat. No.
<i>EpiQuik</i> [™] Total Histone H3 Quantification Kit (Fluorometric)	48 assays 96 assays	P-3063-48 P-3063-96

Available Related Products

	Cat. No.
<i>EpiQuik</i> [™] Global Di-Methyl Histone H3-K4 Quantification Kit	P-3022
<i>EpiQuik</i> [™] Global Di-Methyl Histone H3-K4 Quantification Kit (Fluorometric)	P-3023
<i>EpiQuik</i> [™] Global Mono-Methyl Histone H3-K4 Quantification Kit (Colorimetric)	P-3024
<i>EpiQuik</i> [™] Global Mono-Methyl Histone H3-K4 Quantification Kit (Fluorometric)	P-3025
<i>EpiQuik</i> [™] Global Tri-Methyl Histone H3-K4 Quantification Kit (Colorimetric)	P-3026
<i>EpiQuik</i> [™] Global Tri-Methyl Histone H3-K4 Quantification Kit (Fluorometric)	P-3027
<i>EpiQuik</i> [™] Global Pan-Methyl Histone H3-K4 Quantification Kit (Colorimetric)	P-3028
<i>EpiQuik</i> [™] Global Pan-Methyl Histone H3-K4 Quantification Kit (Fluorometric)	P-3029
<i>EpiQuik</i> [™] Global Histone H3 Quantification Kit (Colorimetric)	P-3062

Need more components? You can also order parts separately by calling 1-877-374-4368 or e-mailing sales@epigentek.com.

